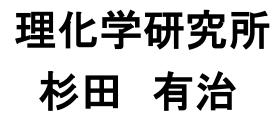




# 細胞環境を考慮した生体分子シ ミュレーションの現状と今後

### PCクラスタシンポジウム 2013/12/13







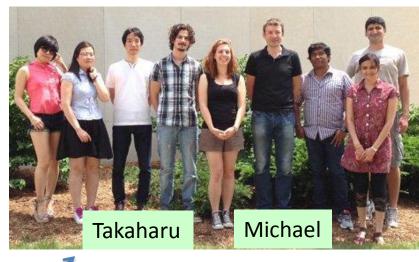
## People who are working for the research

#### Sugita Group in RIKEN (Wako & Kobe)



- **RIKEN Theoretical Molecular Science Laboratory** 
  - Dr. Isseki Yu
  - Dr. Takaharu Mori (Visiting Scientist in MSU)
- RIKEN AICS
  - Dr. Jaewoon Jung
  - Dr. Ryuhei Harada (→ Programming Environment Research Team)

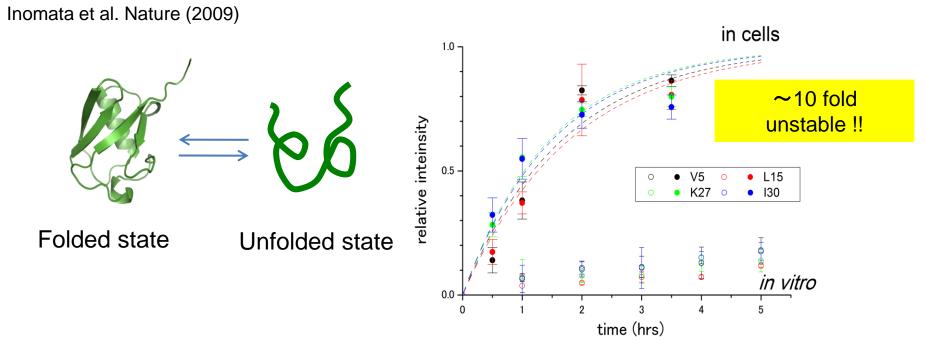
### Feig Group in Michigan State University



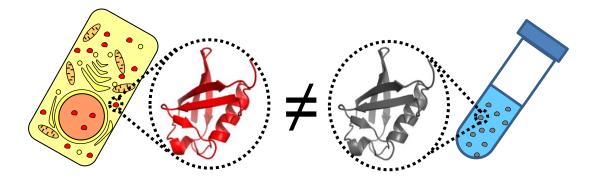
- Michigan State University
  - Prof. Michael Feig
- RIKEN QBiC (NMR experiment)
  - Dr. Takanori Kigawa
  - Dr. Naoya Tochio

## Protein Crowding Simulations and Comparison with NMR experiments

## Why cellular environment is important ?

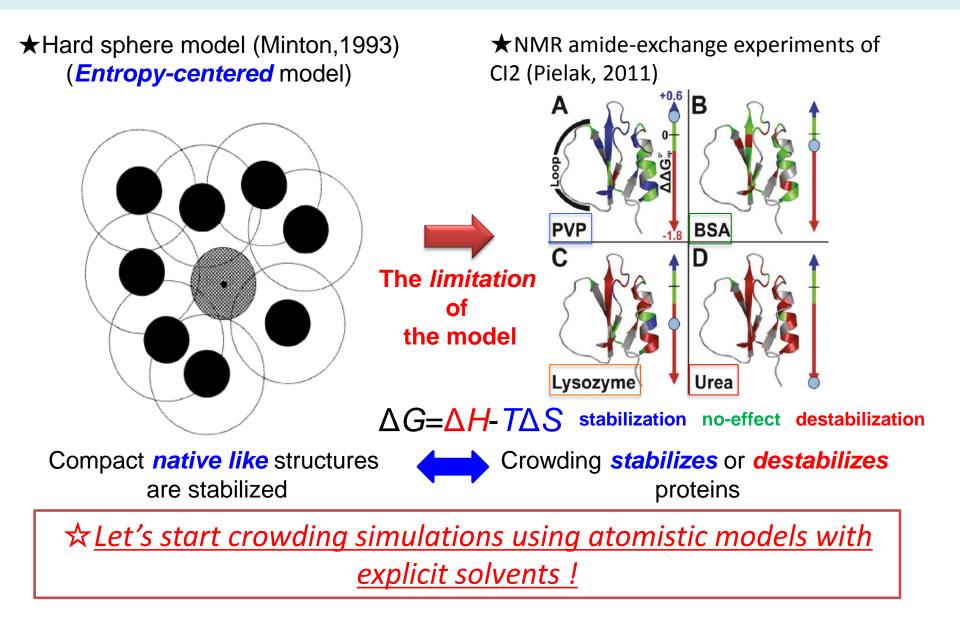


In-cell NMR suggests that conformational stability of ubiquitin in cells is lower than *in vitro*.



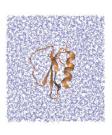
Protein dynamics is different from *in vitro*.

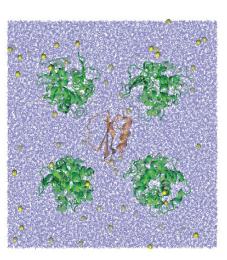
### **Macromolecular Crowding: Theory vs Experiment**

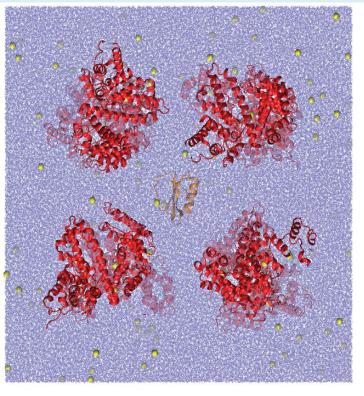


# Protein Crowding Systems for comparing with NMR experiments

Feig, Sugita: J. Phys. Chem B (2012) 116, 599-605







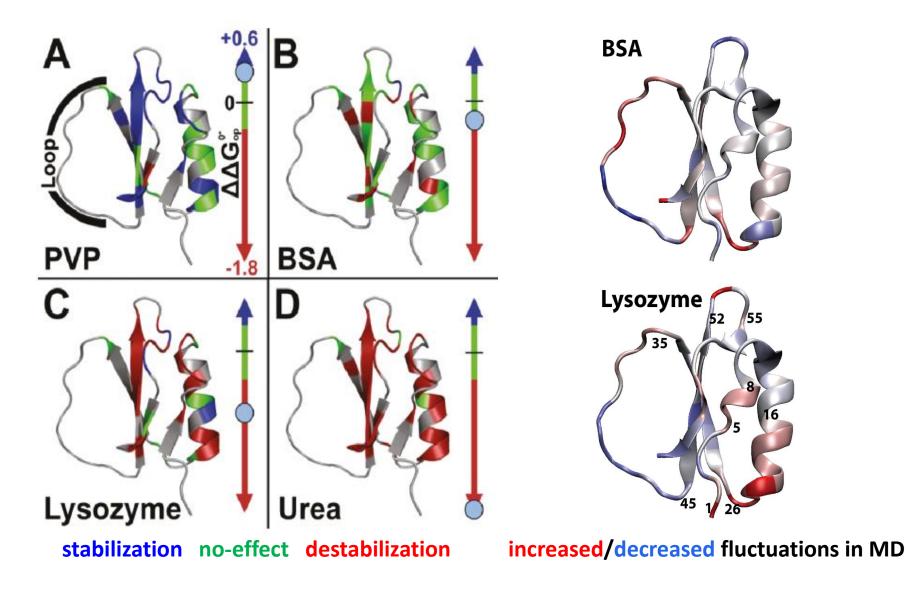
CI2 18K atoms 6K H<sub>2</sub>O infinite dilution Cl2 + 8 lysozymes 184K atoms 56K H<sub>2</sub>O/64 Cl<sup>-</sup> 108 g lysozyme/L 7% vol fraction CI2 + 8 BSAs 835K atoms 253K H<sub>2</sub>O/136 Na<sup>+</sup> 104 g BSA/L solution 7% vol. fraction

~160 ns MD

~250 ns MD

~120 ns MD

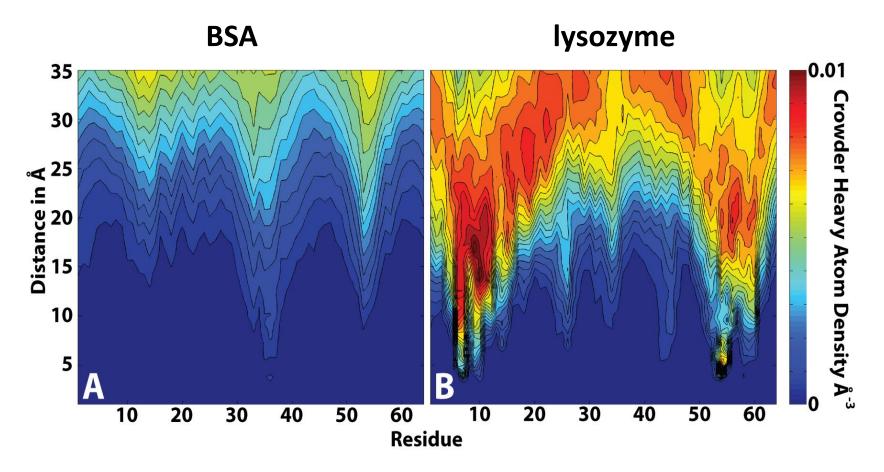
### **Protein Stability in NMR vs Protein Fluctuation in MD**



A. Miklos, M. Sarkar, Y. Wang, G. Pielak: JACS (2011) 133, 7116-7120

Feig, Sugita: J. Phys. Chem B (2012) 116, 599-605

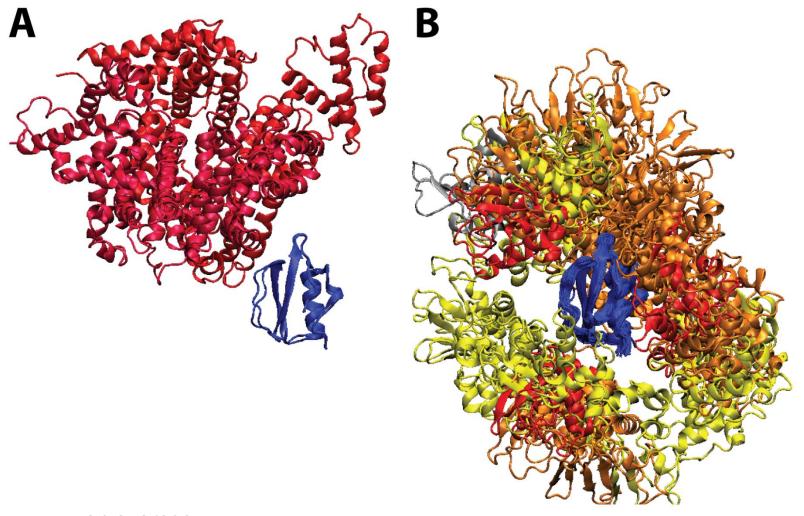
# Interaction between CI2 and crowder proteins (BSA or Lysozyme)



Feig, Sugita: J. Phys. Chem B (2012) 116, 599-605

Interaction between CI2 and lysozyme is stronger than that between CI2 and BSA.

### **CI2 interaction with crowder proteins**



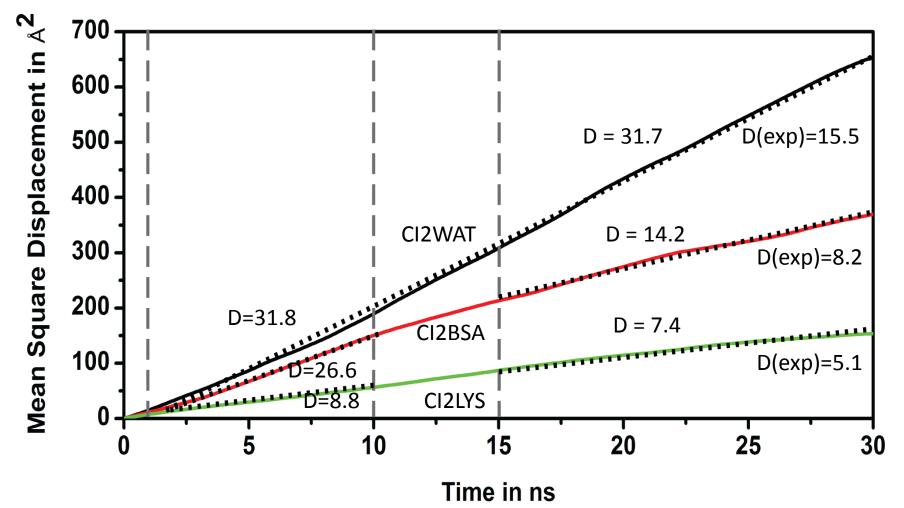
on average 1.1 CI2-BSA contacts

**2.6 CI2-lysozyme contacts** 

Feig, Sugita: J. Phys. Chem B (2012) 116, 599-605

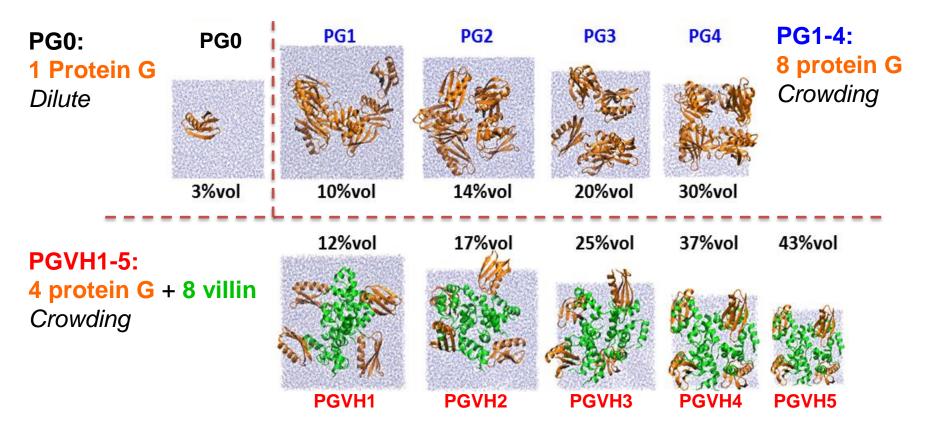
### **Cellular Environment slow down diffusion of proteins**





 $[\mu m^2/s]$  obtained according to Einstein relationship from slope of mean square displacement vs time.

### **Crowding Systems with different concentration of proteins**



Harada, Sugita, Feig, J. Am. Chem. Soc. 2012, 134, 4842-4849

NPT(1bar, 298K, 300ns)

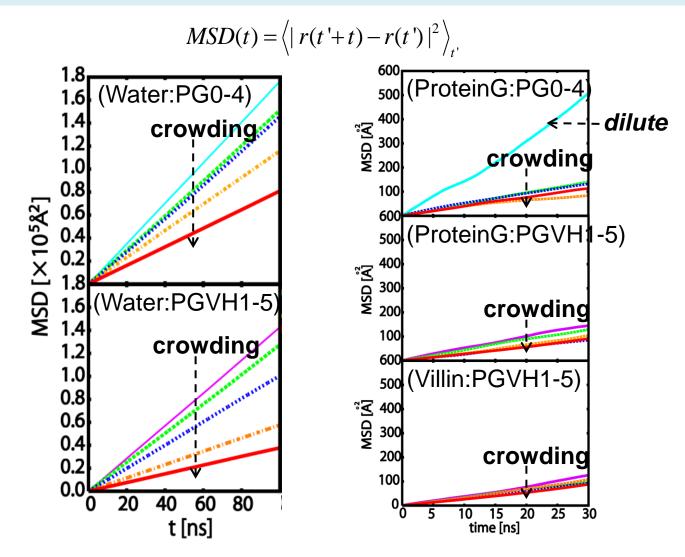
# Available volume for water within the first and second solvation shell vs bulk

#### Harada, Sugita, Feig, J. Am. Chem. Soc. 2012, 134, 4842-4849

System	Protein volume fraction	1 <sup>st</sup> solvation shell (r ≤ 4 Å) [%]	2 <sup>nd</sup> solvation shell (4 Å < r ≤ 7 Å) [%]	Bulk (r >7 Å) [%]
PG1	0.10	22.4	14.4	63.3
PG2	0.14	31.7	20.4	47.9
PG3	0.20	44.0	24.6	31.5
PG4	0.30	65.6	26.7	7.7
PGVH1	0.12	28.1	16.3	55.6
PGVH2	0.17	37.7	19.1	43.3
PGVH3	0.25	53.4	22.2	24.5
PGVH4	0.37	76.2	19.6	4.2
PGVH5	0.43	86.4	13.0	0.6

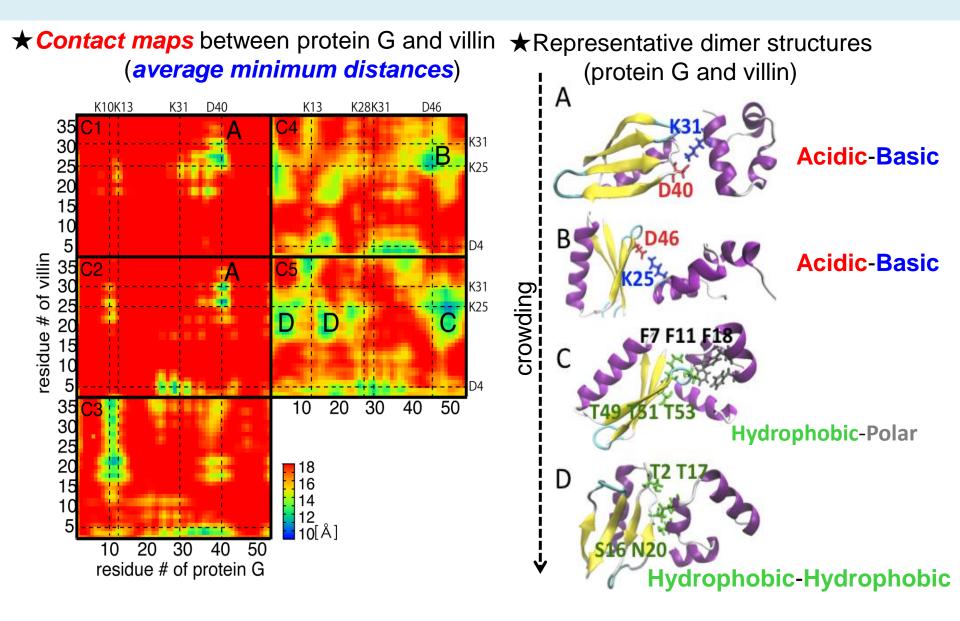
★ Almost no room for bulk water in highly crowded conditions (PG4, PGVH4, PGVH5).

### Diffusion of Water and Protein Molecules in the Crowding Systems



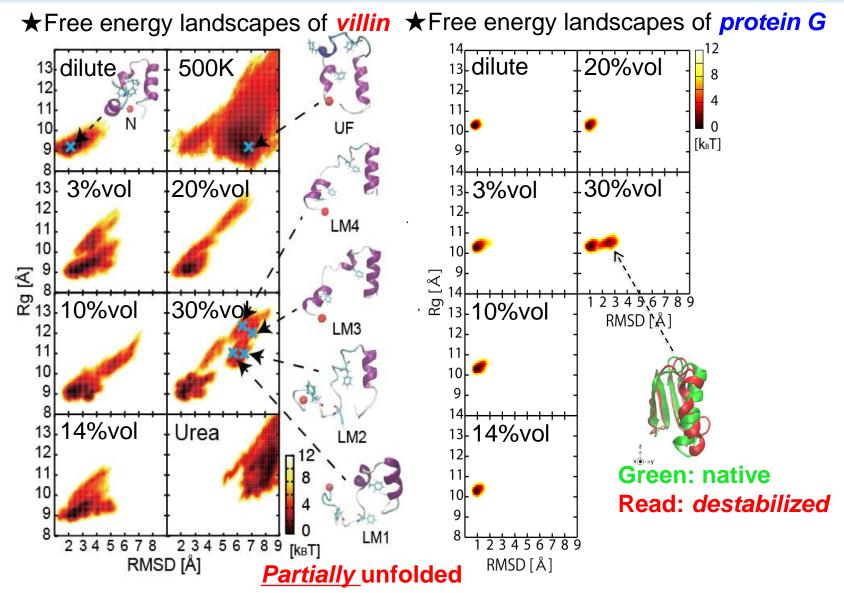
★ In crowded environment, diffusion of water and protein significantly slow down Harada, Sugita, Feig, J. Am. Chem. Soc. 2012, 134, 4842-4849

### **Protein-Protein Interactions in Crowding Systems**



Harada, Tochio, Kigawa, Sugita, Feig, J. Am. Chem. Soc. 2013, 135, 3696-3701.

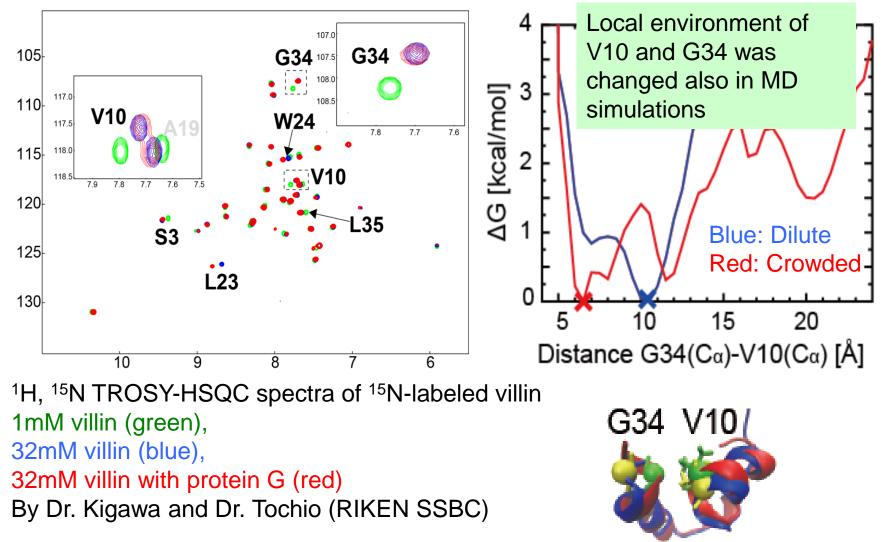
# Conformational Stability of Villin and Protein-G in different crowding systems



Harada, Tochio, Kigawa, Sugita, Feig, *J. Am. Chem. Soc.* 2013, 135, 3696-3701.

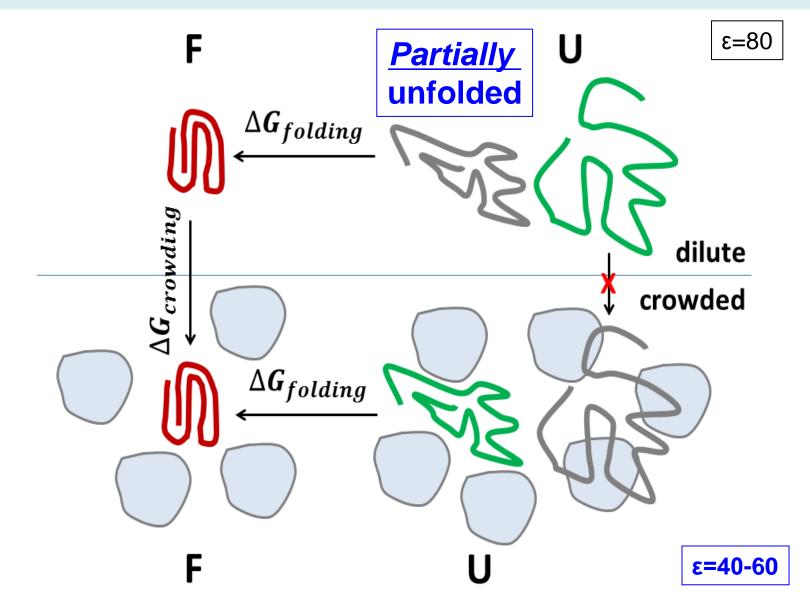
### Comparison of protein crowding systems between MD and NMR

V10 and G34 changes their chemical shift due to crowding



Harada, Tochio, Kigawa, Sugita, Feig, J. Am. Chem. Soc. 2013, 135, 3696-3701.

### New views on macromolecular crowding



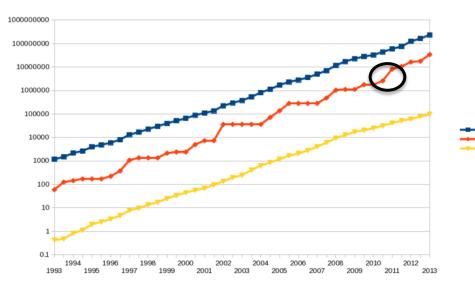
Harada, Tochio, Kigawa, Sugita, Feig, *J. Am. Chem. Soc.* 2013, 135, 3696-3701.

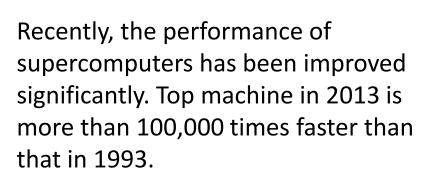
## Development of Highly Parallelized MD Software: GENESIS

**©RIKEN** 

### K computer at RIKEN Advanced Institute for Computational Science (AICS)

### Performance of SuperComputer (from Top500)







### K computer (The 4th fastest computer) Peak Performance: 10.51 PFLOPS # of CPU Cores: 705,024 Total Memory: 1.41PB(16GB per node) Network: Tofu: 6D mesh / Torus

### **GENESIS (**<u>Gen</u>eralized-<u>E</u>nsemble <u>Si</u>mulation <u>Systems</u>)

- 1. Aims at developing efficient and accurate methodologies for free-energy calculations in biological systems
- 2. Efficient Parallelization Suitable for massively parallel computers, in particular, K computer
- 3. Applicability for large scale simulation
- 4. Algorithms coupled with different molecular models such as coarse-grained, all-atom, and hybrid QM/MM
- Generalized ensembles like Replica-Exchange Molecular Dynamics (T-REMD, REUS, MREM, Surface-tension REMD (New! T.Mori et al. JCTC in press.)
- 6. Open Source Code from this December

### **GENESIS (**<u>Gen</u>eralized-<u>E</u>nsemble <u>Si</u>mulation <u>Systems</u>)

### **GENESIS (V1) Development Team**

- Project Leader: Yuji Sugita
- Major Developers: Jaewoon Jung, Takaharu Mori
- Developers: Chigusa Kobayashi, Yasuhiro Matsunaga, Takashi Imai, Takao Yoda (Nagahama Bio Institute), Norio Takase (Isogo Soft)
- Other Contributors: Many members in Sugita Group











Y. Sugita

J. Jung

T. Mori

C. Kobayashi

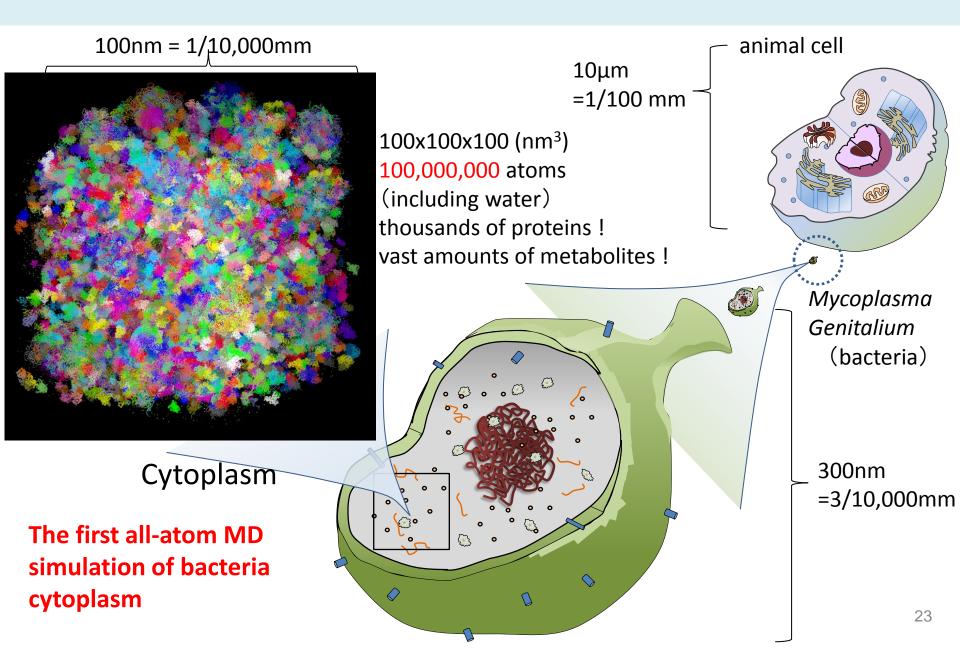
Y. Matsunaga

### New Features of GENESIS (V1)

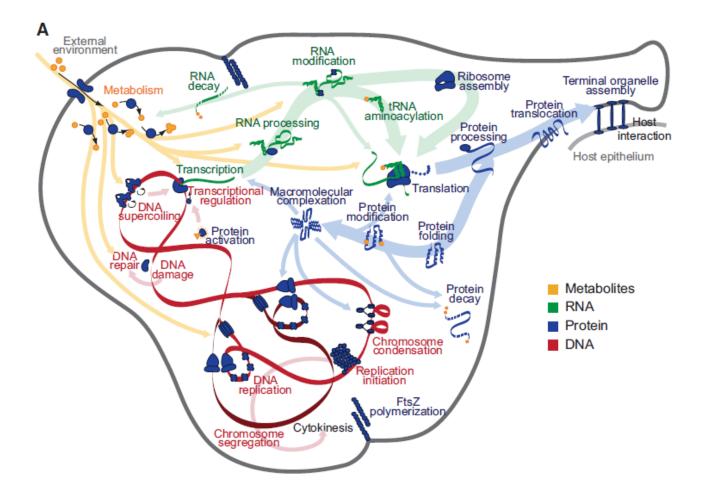
- Inverse Lookup Table Scheme for Nonbonded Interaction
  - J. Jung et al. J.Comp.Chem. 2013, 34, 2412-2420.
- Midpoint Cell Method for Hybrid Parallelization
- Fast 3D-FFT Calculations

## All-atom MD Simulations of Bacterial Cytoplasm on K computer

### **Our Target System for All-atom MD Simulation**

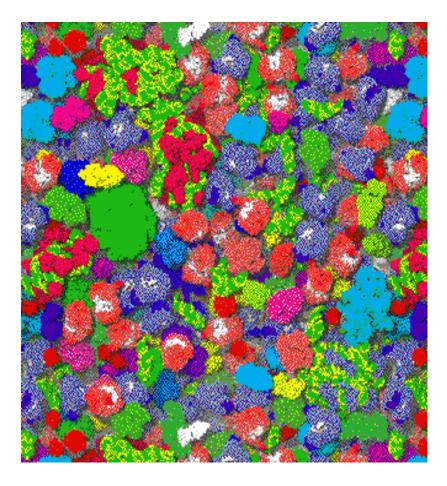


# Mycoplasma genitalium: a bacterium with the smallest known genome



Karr, J.R., Sanghvi, J.C., Macklin, D.N., Gutschow, M.V., Jacobs, J.M., Bolival, B., et al. A Whole-Cell Computational Model Predicts Phenotype from Genotype. Cell. 2012, 150, 389-401.

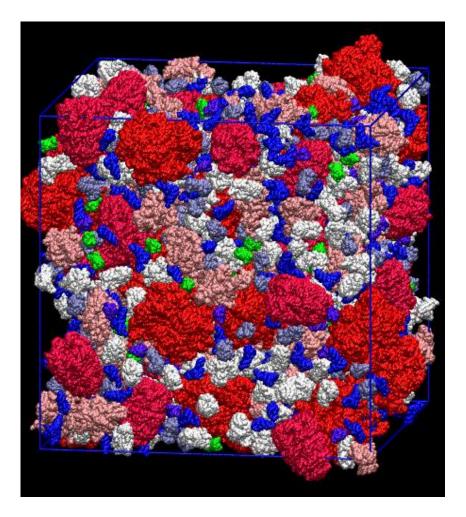
### **Brownian Dynamics Simulations by Elcock et al.**



- PLoS Computational Biology 2010
- 80 X 80 X 80 (nm<sup>3</sup>)
- 6 micro sec.
- 50 selected proteins from E. coli
- No metabolites/solvents
- Effective potential.
- Diffusion coefficients are provided as an input parameters.
- Focus on Thermodynamics (Protein Stability)
- Advanced treatment of molecules (full sampling and full scoring) is required to predict protein stability in crowded conditions.

### **Coarse-grained BD Simulations Without Hydrodynamic Effect**

### **Brownian Dynamics Simulations by Ando and Skolnick**



- PNAS 2010, 107: 18457-18462
- 100 X 100 X 100 (nm<sup>3</sup>).
- A virtual cytoplasmic system of E. coli
- Over 1,000 macromolecules consisting of 15 different macromolecules in Brownian Dynamics (BD) simulations (50 micro sec)
- Focus on the diffusion coefficient
- Hydrodynamic interactions greatly reduce the diffusion coefficient and create collective motions at cellular concentrations.

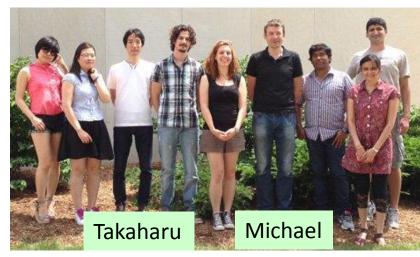
### **BD simulations with hydrodynamic effect on Spherical Model**

### Acknowledgement

#### Sugita Group in RIKEN (Wako & Kobe)



#### Feig Group in Michigan State University



#### Computational Resources

- RIKEN RICC
- K computer
- Research Fund
  - SPIRE Field 1
  - RIKEN Internal funds (QBiC, AICS)

